

Identification of fungi associated with storage corn (zea mays) and estimate fumonisin B1 production by *Fusarium* spp. isolated from these cereals

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Abstract

Zea mays one of the most important grains produced in Babylon province of Iraq incidences of contaminated seeds with toxigenic fungi were investigated in Babylon maize in 2016 from four Locations (centres (of accumulation corn seeds) namely medhateia, Babil, mussaieb and Locally markets, present study included identify the mycoflora of maize in these locations. Mycological analyses showed a predominance of *Fusarium verticillioides* (48.5%), followed by *Aspergillus flavus* (16.5%), *F. proliferatum* (6.5%), *Penicillium* species (5%), *Mucor* species (1.43%) in maize

Sixty strains of *Fusarium verticillioides* and *F. proliferatum* isolated from corn collected from fourth stores in Babylon province, were evaluated for their ability to produce fumonisins B1 (FB1), in Rice culture. Fumonisin levels were determined by high-performance liquid chromatography. All tested strains of *F. verticillioides* and *F. proliferatum* produced fumonisins within a wide range of concentrations, 233-1471 ppb, 176-823 ppb, 134-792 ppb and 236-1271 ppb FB1. The highest mean concentrations of FB1 were 674, 617, 548 and 340 ppb for location (locally markets, medhateia, mussaieb and babil, , respectively) from the four locations of province were observed. Fumonisin mean levels produced by *F. verticillioides* and *F. proliferatum* strains isolated from locally market zone) were significantly ($P < 0.05$) higher than the other three location.

Keyword: *Fusarium*, *Aspergillus penicillium*, Fumonisin, corn

Introduction

spread seed-borne fungi and contamination by mycotoxin producing species in maize [1], is important in evaluate the risk of mycotoxin contamination. A number of fungal species related with maize, mainly belonging to the genera *Fusarium* and *Aspergillus*, and *penicillium* have been reported to produce mycotoxins which cause mycotoxicoses in human and animals [2]. In some maize producing countries, information is available regarding the mycoflora of stored corn cereals, including Argentina [3]; United States, South Africa, Canada [4] and some other countries [5]. The growth of fungi in store is controlled via the following factors such as: (1) composition of nutrients in the seeds (2) humidity and temperature conditions (3) biotic factors as competition or the founding of stored product or damage of insects. Storage fungi are much more frequent in lots infested via stored product insects, because insects generate humidity and separate fungi spores in the commodity [6]. Storage fungi require a relative humidity of at least 65% which is equivalent to an equilibrium moisture content of 13% in cereal grain. They grow at temperatures of between 10 and 40°C [7].

Mycotoxins are secondary metabolic toxic substances which are produced by different fungi. They stay in the stored produce as residues. Mycotoxins can be found in the stored products as soon as 24 h after infestation with fungus. The suitable climatic conditions for the growth of

fungi and the formation of mycotoxins are almost not corresponding and dependent on diverse unidentified factors [8].

All mycotoxins are low-molecular-weight natural products (that is, small molecules) produced as secondary metabolites by filamentous fungi

The fumonisins are a group of structurally associated mycotoxins produced fundamentally by *Fusarium verticillioides* (Sacc.) Nirenberg (for *F. moniliforme* Sheldon) and *F. proliferatum* (Matsushima) Nirenberg, both of which frequently infect Maize [1] crops worldwide [9].

Fumonisin were first isolated in 1988 from cultures of *F. verticillioides* MRC 826, which was firstly gained from corn in a high oesophageal cancer (OC) rate area in the Transkei region of South Africa [10, 11]. *F. verticillioides* and *F. proliferatum* have been isolated from corn worldwide global those that appear well these fungi have been recognized as the highest fumonisin producers, and most isolates of these two species are able to produce fumonisin [12].

In spite of 28 structurally associated fumonisin analogues have been recognised only three, fumonisins B1 (FB1), B2 (FB2) and B3 (FB3) occur at plenty levels. They are frequently appeared in fungal cultures and/or in naturally contaminated corn, which is an important of ingredient human and animal diets in many countries [13]. FB1 generally represents 70–80% of the total fumonisin levels, FB2 accounts for 15–25% and FB3 usually makes up from

3 to 8% when cultured on corn, rice or in liquid medium [12].

Fumonisin causes many diseases on animal such as leukoencephalomalacia (LEM) in horses [14], edema in pigs and liver cancer in Rats [11] also effect on human studies revealed correlation between consumption diet contaminated with *F.verticilloides* or *F.proliferatum* With high of OC [12].

The International Agency for Research on Cancer (IARC) in 2002 (IARC 2002) classified FB1 as possibly carcinogenic to humans (class 2B carcinogen).

F. verticillioides and *F. proliferatum* as the most common seed borne fungi in many country south Africa and in northern Iran.

In present study, corn designed for human and animal consumption in Babylon /Iraq was collected from four main stores of corn creals .

The production of Fumonisin by strains of *F. verticillioides* and *F. proliferatum*, isolated as internal mycoflora from corn seeds, was evaluated. The fumonisin levels produced by these *Fusarium* species were compared between the four stores.

Metrial Methods:

samples of seeds (each ca. 100seed) from each sample were surface sterilized by immersion in (2%)sodium hypochlorite solution in a 100ml Erlenmeyer flask for 1 min, washed twice in steriledistilled water and dried in two layers of sterilised filter papers. (5 per 90 mm plate) on PDA medium containing 500 mg/Lof chloramphenicol to prevent the growth of bacteria [1]. After incubationfor 5–7 days at 25°C in the dark the developing fungal colonies were counted directly and where different fungi were isolated from single seed all the colonies were recorded and different species subcultured onto potato dextrose agar (PDA) [15].

Identification of fungi

Identification of *Fusarium* species subcultures were made on PDA incubated for 7 days at 25°C. Final identifications were made following [4] for [13, 16].

The isolation frequency (Fq), and the incidence of genera and species isolated [17, 18].

were calculated as follows:

Frequency (%) = Number of samples in which a genus/species occurred × 100 / Total number of samples

Incidence (%) = Number of grains infected by a genus/species × 100 / Total number of grains

Ability of Fusarium isolate for production Fumonisin toxin

isolates of *F. verticillioides* and *F. proliferatum* were cultured on rice medium [1] for FB1 detection.

Extraction and Cleanup

FB1 was extracted from the finely ground sample using a protocol developed by Maheshwar et al. [19]. 10g samples were placed into a 250 ml conical flask containing 50 ml of acetonitrile: water (ACN: water, 50+50, v/v) and the flask was covered and shaken for 30 min. 15 ml of the supernatant was filtered through a Whatman No. 4 filter paper. A C18 Sep Pak solid phase extraction (SPE) clean up column was preconditioned by rinsing with 2 ml ACN

followed by 2 ml of 1% aqueous potassium chloride (KCl). The filtered extract (2 ml) and 1% KCl (6 ml) were mixed in a vial and applied to the column [19].

High performance liquid chromatography (HPLC) analysis involved use of ahplc instrument SYKAM for quantification of fumonisin FB1with Fluorescne detector (RF-20A) used for all isolates which be studied.

Results and discussion

Isolation and identification of fungi related corn

A total of 60 fungal isolates were rescued from maize samples collected from the four locations maize Stores in Bbylon provinces of Iraq at 2016. Established onmeans of incidences *Fusarium verticilloides* were the most frequent (48.0%) followed by *Aspergillus flavus* (16.5%), *F.proliferatum* (6.5%), *penicillium*(5%), *Rhizopus* sp.(3.5%) and *Mucor* species (1.43%) (Other genera, were isolated at low frequencies shown in table (1)

%		Isolated fungi
Frequency	Incidence	
1.43*	50	<i>Alternaria alternata</i>
17.85	100	<i>Aspergillus flavus</i>
5.00	100	<i>A niger</i>
1.43	25	<i>A.oryzae</i>
2.14	50	<i>A prasiticus</i>
1.43	25	<i>A. terreus</i>
4.28	75	<i>Fusarium proliferatum</i>
3.57	50	<i>F. solani</i>
48	100	<i>F. verticelloides</i>
4.28	100	<i>Fusarium spp</i>
1.43	25	<i>Mucor spp</i>
5.00	75	<i>Penicillium spp</i>
3.75	50	<i>Rhizoctonia solani</i>
3.56	25	<i>Rhizopus spp</i>
100 %		Total

Every number represent four replicates. *

The spread of *F. verticillioides* as a predominant seed-borne fungus in storage seeds of Babylon stores of, Iraq was contrast to the other studies by Aiyaz et al. [20], Moshiri et al. [21] and countrie s such as South Africa [11] United States [22] Brazil [23] with the sutable climatic conditions of this province to this fungi *F. proliferatum* was the second most predominant *Fusarium* species. Several studies on the distribution of *F. proliferatum* with *F.verticilloides* on corn, rice [24].

HPLC detection of fumonisin in stored corn grains

In the present study ability of producing fumonisin by *Fusarium* strains from four locations was investigated. Detailed results are shown in Table 2 and Fig.1, The 60 strains of *F. verticillioides* and *F.proliferatum* 55strains of *F. verticilloides* 5 strain of *F.proliferatum* all produced FB1

Location	Percentage % incidence	Concentration range (ppb)
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Medhatia	100	233.34-1471
Babil	100	134.5-792.00
Mussaib	100	176.5-821.00
Local markets	100	236.00-1275

There was a high degree of variability in the concs. of fumonisins produced by *F. verticillioides* and *F. proliferatum*. In some cases, large differences in fumonisin production can be observed between strains recovered

from the same location or from the same store. The *F. verticillioides* culture material had FB1 levels ranging from 233.34–1471ppb, 134–792 ppb, 176.5–821 ppb and 236–1275ppb respectively [13, 25].

A further analysis of general mean for the mycotoxins detected by hplc in four location in Babylon province (Figure 1), shows that there was a spread of Fumonisin (B1) in stored corn cereals

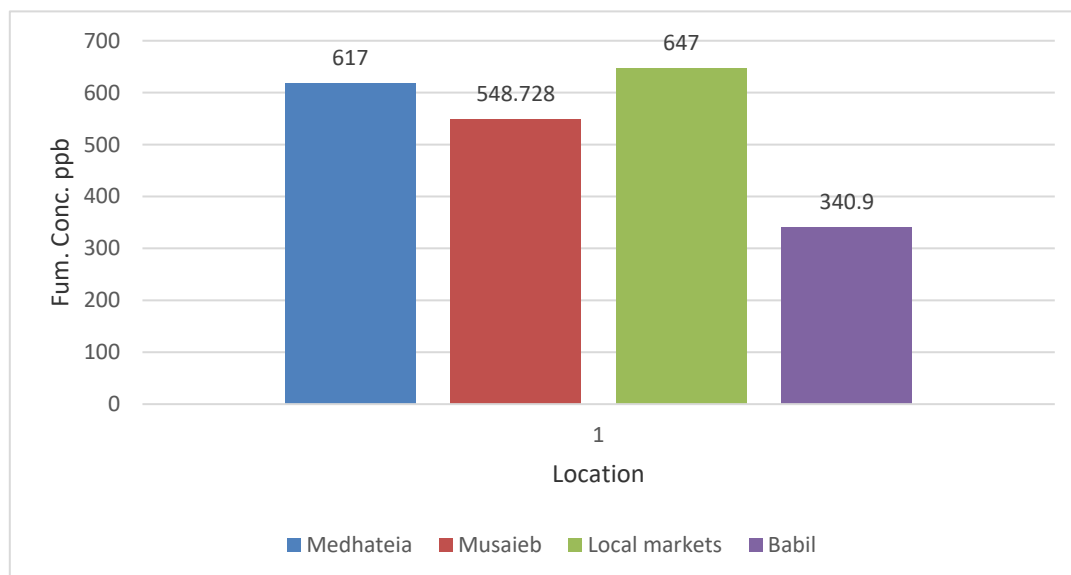


Figure 1. Comparison of Fumonisin contamination between four stores for seed corn

The results explain highest mean levels of FB1 were produced from location (locally markets) i.e. 674ppb and lowest levels from Babil location i.e. 340.93. Our results are in contrast with other reports that many strains of *F. verticillioides* and *F. proliferatum* isolated from corn and other widely differs from different locations that's found in, i.e. North America, Africa, Asia and Australia, have a high potential for fumonisin production [10, 26]. The production of fumonisins by different strains of *F. verticillioides* varies widely.

Commonly, environment conditions such as high temperatures and moisture, unseasonal rains during harvest, and insect damages lead to fungal proliferation and production of mycotoxins. Poor harvesting practices, unsuitable storage and less optimal conditions during transport and marketing can also engage to fungal growth and increase the risk of mycotoxin production [27].

References

- Ishii K, Sawano M, Ueno Y, Tsunoda H. Distribution of zearalenone-producing *Fusarium* species in Japan. *Applied microbiology*. 1974;27(4):625-8. <https://doi.org/10.1128/am.27.4.625-628.1974>
- Marasas WF. Fumonisin: their implications for human and animal health. *Natural toxins*. 1995;3(4):193-8. <https://doi.org/10.1002/nt.2620030405>
- González H, Resnik S, Boca R, Marasas W. Mycoflora of Argentinian corn harvested in the main production area in 1990. *Mycopathologia*. 1995;130(1):29-36. <https://doi.org/10.1007/BF01104346>
- Nelson PE, Marasas W, Toussoun T. *Fusarium species an illustrated manual for identification*. The Pennsylvania State University, 1983. Available from: <https://www.psupress.org/books/titles/0-271-00349-9.html>
- Moubasher A, Elnaghy M, Abdel-Hafez S. Studies on the fungus flora of three grains in Egypt. *Mycopathologia et Mycologia applicata*. 1972;47(3):261-74. <https://doi.org/10.1007/BF02051664>
- Pessu P, Agoda S, Isong I, Adekalu O, Echendu M, Falade T. Fungi and mycotoxins in stored foods. *African Journal of Microbiology Research*. 2011;5(25):4373-82. <https://doi.org/10.5897/AJMR11.487>
- Multon JL. *Preservation and storage of grains, seeds, and their by-products: cereals, oilseeds, pulses, and animal feed*. Lavoisier Pub., 1988. Available from: <https://www.cabdirect.org/cabdirect/abstract/19896768989>
- Anon A, I. *Mycotoxins: economic and health risks*. Council for agricultural science and technology, 1989. Available from: <https://www.cast-science.org/publication/mycotoxins-economic-and-health-risks/>
- Jackson LS, DeVries JW, Bullerman LB. *Advances in Experimental Medicine and Biology*. Plenum Press, 1967.
- Gelderblom W, Jaskiewicz K, Marasas W, Thiel P, Horak R, Vlegaar R, Kriek N. Fumonisin--novel mycotoxins with cancer-promoting activity produced by *Fusarium moniliforme*. *Applied and environmental microbiology*. 1988;54(7):1806-11. <https://doi.org/10.1128/aem.54.7.1806-1811.1988>
- Bezuidenhout SC, Gelderblom WC, Gorst-Allman CP, Horak RM, Marasas WF, Spiteller G, Vlegaar R. Structure elucidation of the fumonisins, mycotoxins from *Fusarium moniliforme*. *Journal of the Chemical Society, Chemical Communications*. 1988(11):743-5.

<https://doi.org/10.1039/C39880000743>

12. Rheeder JP, Marasas WF, Vismer HF. Production of fumonisin analogs by *Fusarium* species. *Applied and environmental microbiology*. 2002;68(5):2101-5. <https://doi.org/10.1128/AEM.68.5.2101-2105.2002>
13. Shephard GS, Thiel PG, Stockenström S, Sydenham EW. Worldwide survey of fumonisin contamination of corn and corn-based products. *Journal of AOAC International*. 1996;79(3):671-87. <https://doi.org/10.1093/jaoac/79.3.671>
14. Kellerman TS, Marasas WFO, Thiel P, Gelderblom W, Cawood M, Coetzer JA. Leukoencephalomalacia in two horses induced by oral dosing of fumonisin B₁. 1990. Available from: <http://hdl.handle.net/2263/41861>
15. Ghiasian SA, Rezayat SM, Kord-Bacheh P, Maghsood AH, Yazdanpanah H, Shephard GS, Westhuizen Lvd, Vismer HF, Marasas WF. Fumonisin production by *Fusarium* species isolated from freshly harvested corn in Iran. *Mycopathologia*. 2005;159(1):31-40. <https://doi.org/10.1007/s11046-004-3899-5>
16. Booth C. The genus *Fusarium*. The genus *Fusarium*. 1971. Available from: <https://www.cabdirect.org/cabdirect/abstract/19710503430>
17. Booth T, Gorrie S, Muhsin TM. Life strategies among fungal assemblages on *Salicornia europaea* aggregate. *Mycologia*. 1988;80(2):176-91. <https://doi.org/10.1080/00275514.1988.12025519>
18. Rajasinghe M, Abeywickrama K, Jayasekera R. Aflatoxigenic *Aspergillus flavus* and Aflatoxin Formation in Selected Spices during Storage. *Tropical Agricultural Research and Extension*. 2010;12(1).
19. Maheshwar PK, Janardhana GR. Natural Occurrence of Toxigenic *Fusarium proliferatum* on Paddy (*Oryza sativa* L.) in Karnataka, India. *Trop Life Sci Res*. 2010;21(1):1-10. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3819065/>
20. Aiyaz M, Divakara ST, Nayaka SC, Hariprasad P, Niranjana SR. Application of beneficial rhizospheric microbes for the mitigation of seed-borne mycotoxigenic fungal infection and mycotoxins in maize. *Biocontrol Science and Technology*. 2015;25(10):1105-19. <https://doi.org/10.1080/09583157.2015.1020760>
21. Moshiri M, Hamid F, Etemad L. Ricin toxicity: Clinical and molecular aspects. *Reports of biochemistry & molecular biology*. 2016;4(2):60. Available from: <https://pubmed.ncbi.nlm.nih.gov/27536698>
22. Zummo N. Sweet sorghum varieties resistant to anthracnose in USA are highly susceptible to the diseases in Brazil. In: *Sorghum and Millets Diseases: A Second World Review*. ICRISAT, Patancheru (AP), India, 1992. p. 289.
23. Orsi RB, Corrêa B, Possi CR, Schammas EA, Nogueira JR, Dias SM, Malozzi MA. Mycoflora and occurrence of fumonisins in freshly harvested and stored hybrid maize. *Journal of Stored Products Research*. 2000;36(1):75-87. [https://doi.org/10.1016/S0022-474X\(99\)00029-6](https://doi.org/10.1016/S0022-474X(99)00029-6)
24. Bailly JD, Querin A, Tardieu D, Guerre P. Production and purification of fumonisins from a highly toxigenic *Fusarium verticilloides* strain. *Revue de médecine vétérinaire*. 2005;1(11):547-54. Available from: <http://www.revmedvet.com/artdes-fr.php?id=1375>
25. Ung-Soo L, Myong-Yur L, Kwang-Sop S, Yun-Sik M, Chae-Min C, Ueno Y. Production of fumonisin B1 and B2 by *Fusarium moniliforme* isolated from Korean corn kernels for feed. *Mycotoxin Research*. 1994;10(2):67-72.
26. Musser SM, Plattner RD. Fumonisin composition in cultures of *Fusarium moniliforme*, *Fusarium proliferatum*, and *Fusarium nygami*. *Journal of Agricultural and Food Chemistry*. 1997;45(4):1169-73. <https://doi.org/10.1021/jf960663t>
27. Bereka T, Kuyu C, Tolera K, Addis E. Current postharvest practices and aflatoxin contamination awareness amongst maize producers in Jimma Zone, Southwest of Ethiopia. *World Mycotoxin Journal*. 2022;15(1):35-43. <https://doi.org/10.3920/WMJ2020.2642>