

Argyreia Nervosa Attenuates Insulin Resistance in Diabetic Gastrocnemius Muscle in Experimental Diabetic Rats: Role of Nrf2/Keap-1 Signaling Pathway

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Abstract

Introduction:

Diabetes interferes with the body's ability to metabolize sugar for energy. By far the most devastating aspect of diabetes is the long term complications it causes. *Argyreia nervosa* is a large climber known for its pharmacological properties which includes anti-microbial, analgesic, anti-inflammatory and various other properties.

Materials and Methods:

Fair groups of rats were taken, six rats in each group. The Group-1 rats were kept normal as such. Group-2 were diabetic rats. Group-3 as diabetic rats treated with *A. nervosa*. Group-4 rats were normal rats treated with *A. nervosa*. Then the animal is sacrificed and the gastrocnemius muscle is isolated. This is further analyzed through gene expression along with Real-time PCR.

Results and conclusion:

The control condition was normal. In diabetic condition Nrf2 was reduced and Keap-1 gene was increased. The present study shows that STZ alters the glucose metabolism which was treated by *A. nervosa*.

Keywords: Insulin resistance, diabetes, novel method, gastrocnemius muscle, gene expression analysis, glucose metabolism, innovative technique.

1. INTRODUCTION

Argyreia nervosa, belonging to the family convolvulaceae is commonly called as elephant creeper or woolly morning glory. This plant grows near edges of lakes and river banks. The leaves of *Argyreia nervosa*

contain β -sitosterol, 1-triacontanol and quercetin. Traditionally it is widely used for its antiviral, anti-bacterial, anti-fungal and anti-inflammatory properties (1).

Diabetes interferes with the body's ability to metabolize sugar for energy. The most devastating aspect of diabetes is the long term complications it can cause. The prevalence of diabetes in India has risen from 7.1% in 2009 to 8.9% in 2019. The most common type of diabetes is type 2 diabetes (T2D). T2D results from the inability of the body to properly use insulin (2). It is also associated with body weight and also concerned for physical inactivity (Kubota et al; 2017).

Nuclear factor erythroid 2-related factor 2 (NRF2) is a transcription factor that regulates the cellular defense against toxic and oxidative insulin. (3) Keap-1 is a gene encoding protein containing KELCH-1 like domains. Thus the present study that streptozotocin can alter the glucose metabolism and assess the modulatory effects of repeated administration of *A. nervosa* to model T2D characteristics in Wistar rats. (4). Our team has extensive knowledge and research experience that has translated into high quality publications (5–14)(15–24)). The aim of this study is to analyze *A. nervosa* attenuates insulin resistance in diabetic gastrocnemius muscle in operational diabetic rats: role of NaF1/Keap-1 Signaling pathway.

2. MATERIALS AND METHODS

Chemicals Used:

The entire chemicals and reagents used in this research were of the molecular and analytical grade acquired from Sigma Chemical Company, and Sisco Research Laboratories (Mumbai, India).

Plant collection:

The species will be verified at Anna Siddha Hospital in Chennai, Tamil Nadu, using *Argyrea nervosa* root powder obtained from a pharmacy.

Extract preparation

The roots of *Argyrea nervosa* powder were Soxhlet extracted with 70% ethanol. The

extract was then filtered with Whatman no. 1 filter paper and the solvent evaporated at reduced pressure by using a Rotary evaporator apparatus to get a viscous mass, which was then stored at 4°C until used.

Animals

Animals were maintained as per the National Guidelines and Protocols approved by the Institutional Animal Ethics committee

(BRULAC/SDCH/SIMATS/IAEC/04-2022/109). Healthy adult male Wistar albino rats of Wistar strain (150–180 days old weighing 180–200 g) were used in this study and maintained in clean polypropylene cages at the Biomedical Research Unit and Lab Animal Center (BRULAC), Saveetha Dental College & Hospitals, Saveetha Institute of Medical & Technical Sciences, Chennai – 600 077, Tamil Nadu, India, under specific humidity ($65 \pm 5\%$) and temperature ($21 \pm 2^\circ$) with constant 12 h light and 12 h dark schedule. The standard pellet diet (Lipton India, Mumbai, India) was provided with clean drinking water in ad libitum.

STZ induction

Diabetes was induced in rats by a single intraperitoneal administration of STZ (55 mg/kg) dissolved in 0.1 M citrate buffer, pH 4.5. 48 hours later, blood samples were collected and glucose levels were estimated to confirm the development of diabetes. The rats that showed hyperglycemia (blood glucose level > 250 mg/dl) were selected for experimental study (Shiv 2010).

Grouping of animals

Animals were grouped into 3 groups of six animals each and treated oral administration for 15 days.

Group I – Normal rats

Group II- diabetic rat

Group III - diabetic rat + oral administration of *Argyrea nervosa* 500 mg/kg/day

Group IV - normal rat + oral administration of argyreia nervosa 500 mg/kg/day

Parameters to be studied

Fasting blood glucose (FBG)

After the overnight fasting, the blood glucose was estimated using On-Call Plus blood glucose test strips (ACON Laboratories Inc., USA). From the rat tail tip, the blood was collected and the results were expressed as mg/dl.

Oral glucose tolerance test (OGTT)

For the oral glucose tolerance test, animals fasted overnight. After giving the oral glucose load (10 ml/kg; 50% w/v) blood glucose level was estimated at various time periods (60, 120, and 180 min) by using On-Call Plus blood glucose test strips. Before giving a glucose load, the value of blood glucose is considered as 0 min value. Results were marked as mg/dl.

Fasting serum insulin

Serum insulin was assayed using ultrasensitive rat insulin ELISA kit obtained from Crystal Chem Inc (Illinois, USA). The range of detection is 0.1–64 ng/ml. The percentage cross reactivity of insulin antibody to rat insulin was 100%. The intra assay coefficient of variation was $\leq 10.0\%$ and inter-assay coefficient of variation was $\leq 10.0\%$. Results were expressed as mIU/ml.

Total RNA isolation, cDNA conversion and real-time PCR

Using a TRIR kit (Total RNA Isolation Reagent Invitrogen), total RNA was isolated from control and experimental samples. In brief, to 100 mg of fresh tissue, 1 ml of TRIR was added and homogenized. The content was transferred to a microcentrifuge tube instantly and 0.2 ml of chloroform was added, vortexed for 1 min then kept at 4°C for 5 min. Later, the contents were centrifuged at 12,000 \times g for 15 min at 4°C. The aqueous phase (upper layer) was carefully transferred to a fresh

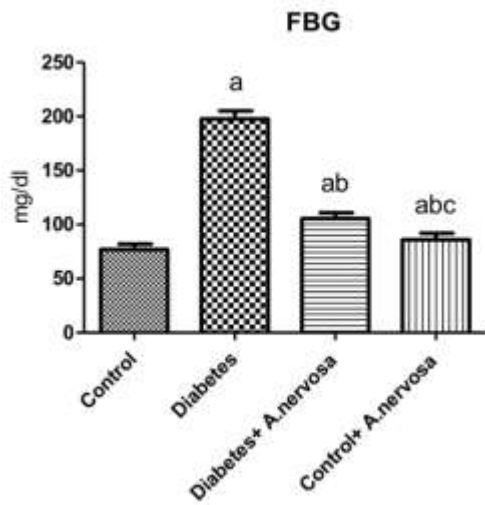
microfuge tube and an equal volume of isopropanol was added, vortexed for 15 S and placed on ice for 10 min. After centrifugation of the content at 12000 \times g for 10 min at 4°C, the supernatant was discarded and RNA pellet was washed with 1 ml of 75% ethanol by the vortex. The isolated RNA was estimated spectrometrically. The RNA concentration was expressed in micrograms (μ g). By using the reverse transcriptase kit from Eurogentec (Seraing, Belgium), complementary DNA (cDNA) was synthesized from 2 μ g of total RNA as stated in the manufacturer's protocol. To perform real-time PCR, the reaction mixture containing 2x reaction buffer (Takara SyBr green master mix), Forward and reverse primers of the target gene and house-keeping gene, water and β -actin (the primer sequences were listed in Table 1) in total volume of 45 μ l except the cDNA was made, mixed intensively and spun down.

In individual PCR vials, about 5 μ l of control DNA for positive control, 5 μ l of water for negative control and 5 μ l of template cDNA for samples were taken and a reaction mixture (45 μ l) were added. 40 cycles (95°C for 5 min, 95°C for 5 s, 60°C for 20 s and 72°C for 40 s) was set up for the reaction and obtained results were plotted by the PCR machine (Stratagene MX 3000 P, Agilent Technologies, 530 l, Stevens Creek Blvd, Santa Clara CA, 95051) on a graph. Relative quantification was calculated from the melt and amplification curves analysis.

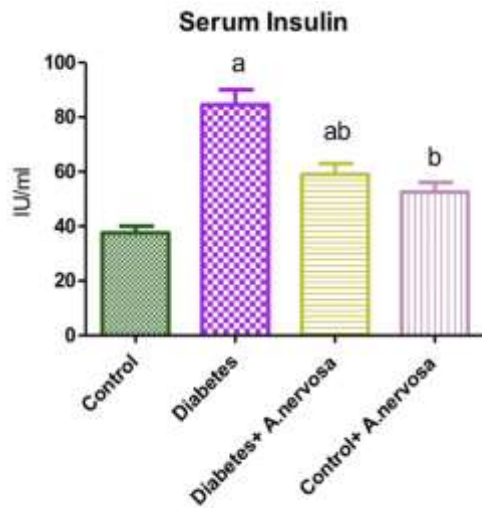
Statistical analysis

The data will be analyzed statistically and ONE-WAY- ANOVA will be used followed by Dencan's multiple range test will be used to check statistical significance among groups. The significance will be considered at the levels of $P < 0.05$.

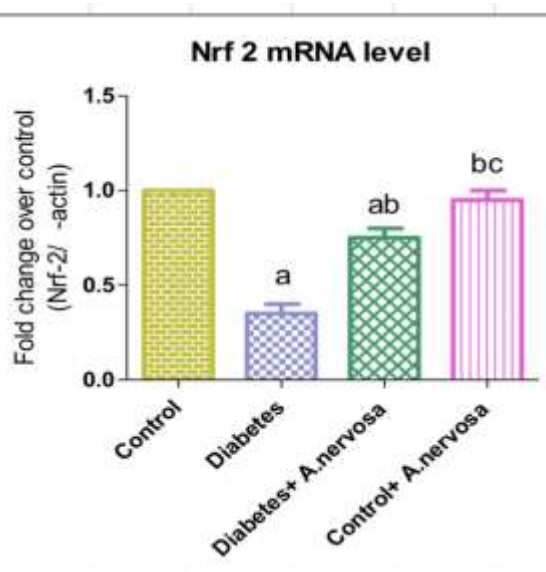
3. RESULTS AND DISCUSSION



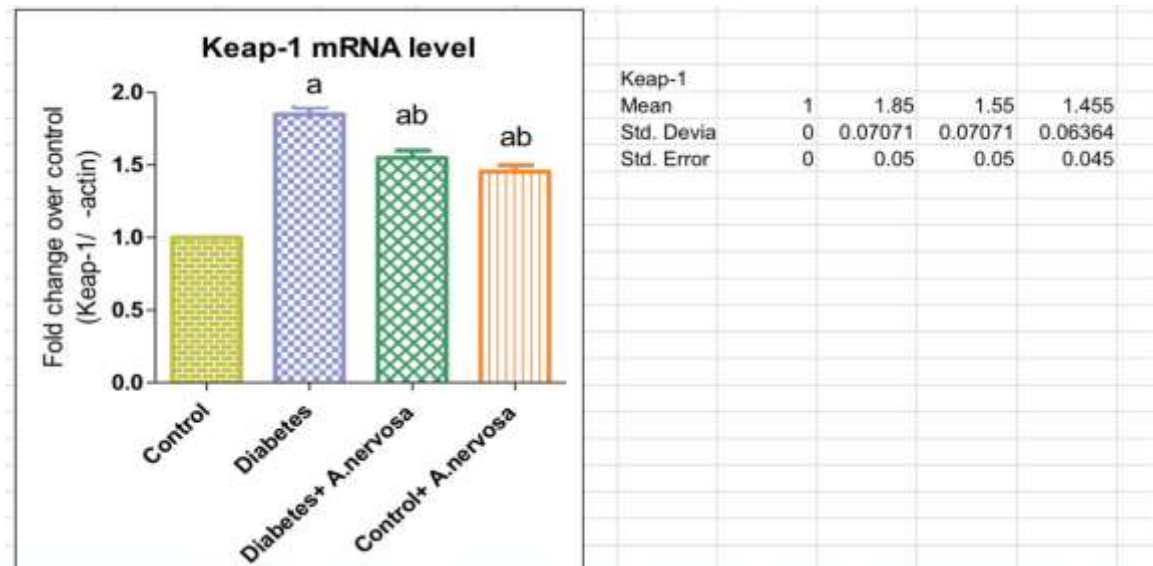
FBG				
Mean	77	197.5	105.5	86
Std. Devia	7.071	10.61	7.778	8.485
Std. Error	5	7.5	5.5	6



Serum insulin				
Mean	37.5	84.5	59	52.5
Std. Devia	3.536	7.778	5.657	4.95
Std. Error	2.5	5.5	4	3.5



Mean	1	0.35	0.75	0.95
Std. Devia	0	0.07071	0.07071	0.07071
Std. Error	0	0.05	0.05	0.05



Insulin resistance promotes impaired fasting blood glucose levels which increases the prevalence of more atherogenic lipid profile molecules. (4,25) The consequences continue to grow into the factors and other cardiovascular diseases depending on the number of components present (26). Preclinical models of experimental rodents are updated.(Engel et al;2019).

By inhibiting the exocytosis of the auxin indolyl acetic acid and by inducing gene expression, flavonoids control plant growth. They also have a variety of additional effects on other biological cells. Their toxicity to mammalian cells, however, is minimal. Many herbal and insect preparations for medical use, such as propolis (bee's glue) and honey, which have been used since ancient times, contain flavonoids as important functional ingredients. Flavonoids should be consumed in amounts of 1-2 g per day with typical meals, particularly fruit and vegetables. Due to their demonstrated capacity to inhibit certain enzymes, mimic some hormones and neurotransmitters, and scavenge free radicals, modern licenced physicians are increasingly using pure flavonoids to treat many serious common disorders (27).

The majority of the diabetic mice received treatment with MSD, FSD, and glibenclamide saw a rise in body weight. Comparing the diabetic control group to the normal control, extracts, and glibenclamide treatment groups, the diabetic control group demonstrated a substantial ($p < 0.05$) drop in body weight (28).

The breakdown of carbohydrates into monosaccharides for absorption is carried out by digestive enzymes for carbohydrates, primarily α -amylase and α -glucosidase. For the treatment of non-insulin diabetes, natural substances made from traditional medicinal herbs that could suppress these digestive enzymes would be helpful. It has been demonstrated that *A. nervosa* seed and leaf compositions have powerful anti-inflammatory and anti-diabetic effects. (29)

In the present study, STZ included group shows the increases in FBG and serum insulin when compared to the control group, whereas in the *A.nervosa* treated group the FBG and serum insulin levels were restored to near normal levels.

The mRNA expression levels of NrF2 was decreased and Keap-1 was increased in diabetic condition, whereas in the *A.nervosa* treated group these genes were

regulated compared with the control group. The present study shows that *A.nervosa* has the anti-diabetic activity by regulating NrF2/Keap-1. Our team has extensive knowledge and research experience that has translate into high quality publications ((30), (31), (32), (33), (34), (35,36), (37), (38), (39), (40).

Plants' metabolic processes result in a wide range of organic chemicals. Because they interact with other plants, animals, and bacteria, these secondary metabolites are essential to the plant's own life. Researchers have spent a lot of time studying flavonoids, a class of secondary metabolites found in plants, because they have beneficial qualities like anti-cholinesterase action, anti-inflammatory activity, radical scavenging activity, etc. As phytoalexins, signal molecules, allopathic substances, detoxifying agents, and antimicrobial defense compounds, they also play a significant role. As they disrupt nucleic acids and proteins, these flavonoids are also claimed to have insecticidal and antibacterial properties. They are crucial substances for creating pesticides for the agricultural sector (41).

Additionally, Lipinski's rule of five for flavonoids was examined. Myricetin and vitexin showed promising results despite breaking just one of Lipinski's rules. The examined flavonoids can be regarded as strong candidates in the process of developing drugs to combat hyperglycemia, despite the fact that several parameters were not reached by them (41,42).

Rats fed a cafeteria meal were used as test subjects for the effects of the ethanolic extract of *A. speciosa* root. The extract demonstrated a reduction in obesity in experimental mice by considerably lowering the serum levels of leptin, total cholesterol, low density lipoprotein, and triglycerides.(28,43).

4. CONCLUSION

The present study shows that streptozotocin alters the glucose metabolism in rats which were treated by *A.nervosa*.In further studies, molecules or genes related to insulin resistance need to be studied to prove *A.nervosa* as a potential anti-diabetic agent.

Conflict of Interest:

The authors hereby declare that there is no conflict of interest in this study.

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Author Contribution:

A) Jeshurun Jegan - contributed in designing the study, execution of the project, statistical analysis, manuscript drafting.

B) Dr. Selvaraj - contributed in designing the study, execution of the project, statistical analysis, manuscript drafting.

C) Dr.V.Vishnupriya - contributed in study design, guiding the research work, manuscript correction.

D) Dr. Gayathri R - study design, statistical analysis, manuscript proofreading and correction.

E) Dr. Kavitha S - study design, statistical analysis, manuscript proofreading and correction.

5. REFERENCES

1. Singhal AK, Gupta H, Bhati VS. Wound healing activity of *Argyrea nervosa* leaves extract. *International Journal of Applied and Basic Medical Research*. 2011;1(1):36.
2. Gupta G, Wadhwa R, Pandey P, Singh SK, Gulati M, Sajita S, et al. Obesity and Diabetes: Pathophysiology of Obesity-Induced Hyperglycemia and Insulin Resistance. *Pathophysiology of Obesity-Induced Health Complications*. 2020;81–97.
3. He F, Ru X, Wen T. NRF2, a Transcription Factor for Stress Response and Beyond. *Int J Mol Sci*. 2020 Jul 6;21(13):4777.
4. Wilson CJ, Chang M, Karttunen M, Choy WY. KEAP1 Cancer Mutants: A Large-Scale Molecular Dynamics Study of Protein Stability. *Int J Mol Sci*. 2021 May 20;22(10):5408.
5. Sathivel A, Raghavendran HRB, Srinivasan P, Devaki T. Anti-peroxidative and anti-hyperlipidemic nature of *Ulva lactuca* crude polysaccharide on D-galactosamine induced hepatitis in rats. *Food Chem Toxicol*. 2008 Oct;46(10):3262–7.
6. Sekar D, Lakshmanan G, Mani P, Biruntha M. Methylation-dependent circulating microRNA 510 in preeclampsia patients. *Hypertens Res*. 2019 Oct;42(10):1647–8.
7. Rajeshkumar S, Menon S, Venkat Kumar S, Tambuwala MM, Bakshi HA, Mehta M, et al. Antibacterial and antioxidant potential of biosynthesized copper nanoparticles mediated through *Cissus arnotiana* plant extract. *J Photochem Photobiol B*. 2019 Aug;197:111531.
8. Lakshmi T, Krishnan V, Rajendran R, Madhusudhanan N. *Azadirachta indica*: A herbal panacea in dentistry - An update. *Pharmacogn Rev*. 2015 Jan;9(17):41–4.
9. Felicita AS, Chandrasekar S, Shanthasundari KK. Determination of craniofacial relation among the subethnic Indian population: a modified approach - (Sagittal relation). *Indian J Dent Res*. 2012 May;23(3):305–12.
10. Thejeswar EP, Thenmozhi MS. Educational research-iPad system vs textbook system. *J Adv Pharm Technol Res*. 2015;8(8):1158.
11. Saravanan A, Senthil Kumar P, Jeevanantham S, Karishma S, Tajsabreen B, Yaashikaa PR, et al. Effective water/wastewater treatment methodologies for toxic pollutants removal: Processes and applications towards sustainable development. *Chemosphere*. 2021 Oct;280:130595.
12. Menon A, Thenmozhi MS. Correlation between thyroid function and obesity. *J Adv Pharm Technol Res*. 2016 Oct;9(10):1568.
13. Sahu D, Kannan GM, Vijayaraghavan R. Size-dependent effect of zinc oxide on toxicity and inflammatory potential of human monocytes. *J Toxicol Environ Health A*. 2014;77(4):177–91.
14. Wang Y, Zhang Y, Guo Y, Lu J, Veeraraghavan VP, Mohan SK, et al. Synthesis of Zinc oxide

- nanoparticles from *Marsdenia tenacissima* inhibits the cell proliferation and induces apoptosis in laryngeal cancer cells (Hep-2). *J Photochem Photobiol B*. 2019 Dec;201:111624.
15. Wadhwa R, Paudel KR, Chin LH, Hon CM, Madheswaran T, Gupta G, et al. Anti-inflammatory and anticancer activities of Naringenin-loaded liquid crystalline nanoparticles in vitro. *J Food Biochem*. 2021 Jan;45(1):e13572.
 16. Reddy P, Krithikadatta J, Srinivasan V, Raghu S, Velumurugan N. Dental Caries Profile and Associated Risk Factors Among Adolescent School Children in an Urban South-Indian City. *Oral Health Prev Dent*. 2020 Apr 1;18(1):379–86.
 17. Eapen BV, Baig MF, Avinash S. An Assessment of the Incidence of Prolonged Postoperative Bleeding After Dental Extraction Among Patients on Uninterrupted Low Dose Aspirin Therapy and to Evaluate the Need to Stop Such Medication Prior to Dental Extractions. *J Maxillofac Oral Surg*. 2017 Mar;16(1):48–52.
 18. Devarajan Y, Nagappan B, Choubey G, Vellaiyan S, Mehar K. Renewable Pathway and Twin Fueling Approach on Ignition Analysis of a Dual-Fuelled Compression Ignition Engine. *Energy Fuels*. 2021 Jun 17;35(12):9930–6.
 19. Barabadi H, Mojab F, Vahidi H, Marashi B, Talank N, Hosseini O, et al. Green synthesis, characterization, antibacterial and biofilm inhibitory activity of silver nanoparticles compared to commercial silver nanoparticles [Internet]. Vol. 129, *Inorganic Chemistry Communications*. 2021. p. 108647. Available from: <http://dx.doi.org/10.1016/j.inoche.2021.108647>
 20. Manickam A, Devarasan E, Manogaran G, Priyan MK, Varatharajan R, Hsu CH, et al. Score level based latent fingerprint enhancement and matching using SIFT feature. *Multimed Tools Appl*. 2019 Feb 1;78(3):3065–85.
 21. Subramaniam N, Muthukrishnan A. Oral mucositis and microbial colonization in oral cancer patients undergoing radiotherapy and chemotherapy: A prospective analysis in a tertiary care dental hospital [Internet]. Vol. 10, *Journal of Investigative and Clinical Dentistry*. 2019. Available from: <http://dx.doi.org/10.1111/jicd.12454>
 22. Rohit Singh T, Ezhilarasan D. Ethanolic Extract of *Lagerstroemia Speciosa* (L.) Pers., Induces Apoptosis and Cell Cycle Arrest in HepG2 Cells. *Nutr Cancer*. 2020;72(1):146–56.
 23. Wahab PUA, Abdul Wahab PU, Senthil Nathan P, Madhulaxmi M, Muthusekhar MR, Loong SC, et al. Risk Factors for Post-operative Infection Following Single Piece Osteotomy [Internet]. Vol. 16, *Journal of Maxillofacial and Oral Surgery*. 2017. p. 328–32. Available from: <http://dx.doi.org/10.1007/s12663-016-0983-6>

24. Krishnamurthy A, Sherlin HJ, Ramalingam K, Natesan A, Premkumar P, Ramani P, et al. Glandular odontogenic cyst: report of two cases and review of literature. *Head Neck Pathol.* 2009 Jun;3(2):153–8.
25. Reaven GM. THE INSULIN RESISTANCE SYNDROME: Definition and Dietary Approaches to Treatment [Internet]. [cited 2022 Nov 12]. Available from: <https://www.proquest.com/openview/87d00d25c788a5db2797564e41bb495b/1?pq-origsite=gscholar&cbl=49287>
26. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature.* 2006 Dec 13;444(7121):840–6.
27. Havsteen BH. The biochemistry and medical significance of the flavonoids. *Pharmacol Ther* [Internet]. 2002 Nov [cited 2022 Dec 19];96(2-3). Available from: <https://pubmed.ncbi.nlm.nih.gov/12453566/>
28. Obafemi TO, Akinmoladun AC, Olaleye MT, Agboade SO, Onasanya AA. Antidiabetic potential of methanolic and flavonoid-rich leaf extracts of *Synsepalum dulcificum* in type 2 diabetic rats. *J Ayurveda Integr Med* [Internet]. 2017 Oct [cited 2022 Dec 19];8(4). Available from: <https://pubmed.ncbi.nlm.nih.gov/28917550/>
29. Saratale GD, Saratale RG, Benelli G, Kumar G, Pugazhendhi A, Kim DS, et al. Anti-diabetic Potential of Silver Nanoparticles Synthesized with *Argyrea nervosa* Leaf Extract High Synergistic Antibacterial Activity with Standard Antibiotics Against Foodborne Bacteria. *J Cluster Sci.* 2017 Feb 13;28(3):1709–27.
30. Surapaneni KM, Vishnu Priya V, Mallika J. Effect of pioglitazone, quercetin, and hydroxy citric acid on vascular endothelial growth factor messenger RNA (VEGF mRNA) expression in experimentally induced nonalcoholic steatohepatitis (NASH). *Turk J Med Sci.* 2015;45(3):542–6.
31. Zhao Y, Dang M, Zhang W, Lei Y, Ramesh T, Priya Veeraraghavan V, et al. Neuroprotective effects of Syringic acid against aluminium chloride induced oxidative stress mediated neuroinflammation in rat model of Alzheimer's disease. *J Funct Foods.* 2020 Aug 1;71:104009.
32. Dave PH, Vishnupriya V, Gayathri R. Herbal remedies for anxiety and depression-a review. *Research Journal of Pharmacy and Technology.* 2016;9(8):1253–6.
33. Balaji V, Priya VV, Gayathri R. Awareness of risk factors for obesity among College students in Tamil Nadu: A Questionnaire based study. *Research Journal of Pharmacy and Technology; Raipur.* 2017 May;10(5):1367–9.
34. Manohar J, Gayathri R, Vishnupriya V. Tenderisation of meat using bromelain from pineapple extract. *Int J Pharm Sci Rev Res.* 2016;39(1):81–5.

35. Mohan SK, Veeraraghavan VP, Jainu M. Effect of pioglitazone, quercetin and hydroxy citric acid on extracellular matrix components in experimentally induced non-alcoholic steatohepatitis. *Iran J Basic Med Sci.* 2015 Aug;18(8):832–6.
36. Surapaneni KM, Priya VV, Mallika J. Pioglitazone, quercetin and hydroxy citric acid effect on cytochrome P450 2E1 (CYP2E1) enzyme levels in experimentally induced non alcoholic steatohepatitis (NASH). *Eur Rev Med Pharmacol Sci.* 2014;18(18):2736–41.
37. Bolla SR, Mohammed Al-Subaie A, Yousuf Al-Jindan R, Papayya Balakrishna J, Kanchi Ravi P, Veeraraghavan VP, et al. In vitro wound healing potency of methanolic leaf extract of *Aristolochia saccata* is possibly mediated by its stimulatory effect on collagen-1 expression. *Heliyon.* 2019 May;5(5):e01648.
38. Selvaraj K, Sivakumar G, Priya Veeraraghavan V, Dandannavar VS, Veeraraghavan GR, Rengasamy G, et al. *Asparagus Racemosus* -A Review [Internet]. *Systematic Reviews.* Available from: <https://www.sysrevpharm.org/articles/asparagus-racemosus--a-review.pdf>
39. Ansari S, Bari A, Ullah R, Mathanmohun M, Veeraraghavan VP, Sun Z. Gold nanoparticles synthesized with *Smilax glabra* rhizome modulates the anti-obesity parameters in high-fat diet and streptozotocin induced obese diabetes rat model. *J Photochem Photobiol B.* 2019 Dec;201:111643.
40. Jayaraman S, Devarajan N, Rajagopal P, Babu S, Ganesan SK, Veeraraghavan VP, et al. β -Sitosterol Circumvents Obesity Induced Inflammation and Insulin Resistance by down-Regulating IKK β /NF- κ B and JNK Signaling Pathway in Adipocytes of Type 2 Diabetic Rats. *Molecules* [Internet]. 2021 Apr 6;26(7). Available from: <http://dx.doi.org/10.3390/molecules26072101>
41. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. *J Nutr Sci.* 2016;5:e47.
42. Kulkarni AA, Kamble AD. Flavonoids from *Argyreia nervosa* (Burm.f.) Bojer: A Ready Arsenal against Pests as Well as Diabetes. *Biology and Life Sciences Forum.* 2020 Dec 1;4(1):56.
43. Kumar S, Alagawadi KR, Raghavendra Rao M. Effect of *Argyreia speciosa* root extract on cafeteria diet-induced obesity in rats. *Indian J Pharmacol.* 2011 Apr;43(2):163.